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The *Arabidopsis* PLAT Domain Protein1 Is Critically Involved in Abiotic Stress Tolerance

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**Abstract**

Despite the completion of the *Arabidopsis* genome sequence, for only a relatively low percentage of the encoded proteins experimental evidence concerning their function is available. Plant proteins that harbour a single PLAT (Polycystin, Lipoxigenase, Alpha-toxin and Triacylglycerol lipase) domain and belong to the PLAT-plant-stress protein family are ubiquitously present in monocot and dicots. However, the function of PLAT-plant-stress proteins is still poorly understood. Therefore, we have assessed the function of the uncharacterised *Arabidopsis* PLAT-plant-stress family members through a combination of functional genetic and physiological approaches. PLAT1 overexpression conferred increased abiotic stress tolerance, including cold, drought and salt stress, while loss-of-function resulted in opposite effects on abiotic stress tolerance. Strikingly, PLAT1 promoted growth under non-stressed conditions. Abiotic stress treatments induced PLAT1 expression and caused expansion of its expression domain. The ABF/ABRE transcription factors, which are positive mediators of abscisic acid signalling, activate PLAT1 promoter activity in transactivation assays and directly bind to the ABRE elements located in this promotor in electrophoretic mobility shift assays. This suggests that PLAT1 represents a novel downstream target of the abscisic acid signalling pathway. Thus, we showed that PLAT1 critically functions as positive regulator of abiotic stress tolerance, but also is involved in regulating plant growth, and thereby assigned a function to this previously uncharacterised PLAT domain protein. The functional data obtained for PLAT1 support that PLAT-plant-stress proteins in general could be promising targets for improving abiotic stress tolerance without yield penalty.

**Introduction**

The PLAT domain (PS50095; Polycystin-1, Lipoxigenase, Alpha-toxin and Triacylglycerol lipase) forms a β-sandwich composed of two sheets of four strands each and is an intracellular domain. It occurs in a variety of membrane or lipid associated proteins that are multi-domain proteins, but also in proteins harbouring either a single PLAT domain or repeats [1–5]. Because of its similarity to the C2 domain, the PLAT domain was proposed to function in protein-protein interactions as well as protein-membrane interactions [2,4,6]. Indeed, the PLAT domain of *Caenorhabditis elegans* polycystin LOV-1 and human polycystin-1 interact with ATP-2, an ATP synthase F1 subunit [7], while the human polycystin-1L2 interacts with different types of G-proteins [8]. Importantly, the association with membranes is essential for the proper function of PLAT domain proteins [2,7]. Further, the membrane targeted 11R-Lipoxigenase from *Germinopsis fruticosa* was shown to bind calcium, required to induce its activity [9]. The PLAT domain regulates the catalytic activity in multi-domain proteins, but also in proteins interacting with the PLAT domain [2,6], and was shown to regulate substrate specificity [10]. Whereas substantial experimental data on PLAT domain proteins is available for the animal field, PLAT domain proteins from plants were only poorly studied, despite the fact that genes encoding PLAT domain proteins were isolated from several plant species [3,4,11–16].

Transgenic approaches to improve abiotic stress tolerance often resulted in yield penalties under optimal growth conditions [17,19], while only few studies reported an associated improved plant growth [4,19]. Interestingly, one of these studies addressed...
the so far only studied PLAT-plant-stress protein CaTIN1 from *Capsicum annuum*, however this protein was only studied using heterologous expression in tobacco [4]. Because both gain-of-function and antisense *CaTIN1* expression promoted abiotic and biotic stress tolerance, *CaTIN1* function remained elusive. Proteins that belong to the PLAT-plant-stress protein family (Conserved Domain cd1754) are ubiquitously present in monocot and dicot plant species and harbour a single PLAT domain. Our analyses of the limited in silico expression data available for PLAT-plant-stress proteins indicate transcriptional induction by different abiotic and biotic stimuli. This suggests that PLAT-plant-stress proteins in general could promote tolerance towards stress responses, although no data from functional studies in homologous systems are available for these proteins.

The plant hormone abscisic acid (ABA) regulates different aspects of plant development, such as stomatal aperture [20] and seed germination [21]. ABA production is increased by abiotic stresses and ABA regulated genes strongly overlap with those induced under drought, salinity and less prominently cold stress conditions [22–25]. The ABA stimulated stomatal closure has been shown to serve as primary defence mechanism during the initial phase of biotic stress responses [26,27]. In contrast, ABA mostly negatively regulates the subsequent phases in biotic stress responses by repressing the salicylic acid, ethylene, jasmonic acid and cytokinin signalling pathways [26–29]. ABA-deficient mutants showed enhanced defence responses against *Botrytis cinerea* [30], and virulent bacteria in tomato [31] and *Arabidopsis* [32]. Although these findings suggest that ABA is involved in the crosstalk between abiotic and biotic stress responses, no direct link in the antagonistic interaction between these stresses is available.

Based on comparative genomic analysis, we identified three *Arabidopsis* genes (PLAT1 AT4G39730, PLAT2 AT2G22170 and PLAT3 AT5G65158) that belong to the PLAT-plant-stress subgroup and submitted this annotation to the TAIR database. PLAT1 and PLAT2 are orthologs of *CaTIN1* and *CaTIN1-2*, respectively [4,16]. Our in silico analysis of published experimental data [33] revealed that cold stress induced the expression levels for the PLAT1 ortholog in *Thlaspi arvense*, which is a close relative of *Arabidopsis*. Based on these findings we hypothesised that the *Arabidopsis* members from this PLAT-plant-stress subgroup, similar to *CaTIN1* [4], also promote tolerance towards various stress responses. Here we report on the molecular characterisation and functional analysis of the PLAT-plant-stress subgroup family member AT4G39730 that we designated as *Arabidopsis* PLAT domain protein 1 (PLAT1). We showed that PLAT1 critically functions as positive regulator of abiotic stress tolerance, also promotes plant growth and is a direct target of the ABF transcription factors, which are positive mediators of the ABA signalling pathway [34,35]. The possible practical application to increase abiotic stress tolerance without yield penalty in crop species is discussed.

Materials and Methods

Plasmid construction and plant transformation

Total RNA isolated from *Arabidopsis* (TRIR reagent from Thermo Fischer Scientific, Germany) was reverse-transcribed using the ReverAid™ First strand cDNA synthesis kit (Thermo Fischer Scientific, Germany). Using this cDNA as template, the full-length PLAT1 cDNA was amplified by PCR with the PLAT1-F cDNA and PLAT1-R cDNA primers (Table S1 in File S1). The dexamethasone inducible overexpression construct, 35S::

PLAT1 was created by cloning the PLAT1 cDNA PCR product in the OP shuttle vector pEG647. The resulting OP::PLAT1 cassette was transferred to the binary vector pEG618 harbouring the 35S::LhGR activator component, resulting in 35S::>PLAT1. The PLAT1 rescue/reporter constructs were generated by PCR amplification from genomic DNA with the PLAT1-F and PLAT1-R rescue primers (Table S1 in File S1) to isolate the genomic fragment harbouring 2039 bp PLAT1 promoter sequence, including the 5′-UTR region, and the PLAT1 coding region without stop codon. Subsequently, the venus VFP or GUS reporter proteins were fused in frame to the C-terminus of the PLAT1 protein resulting in PLAT1::PLAT1-YFP and PLAT1::PLAT1-GUS, respectively. The different 35S::ABF1–4 overexpression constructs were created by cloning the ABF1–4 PCR products (ABF1–4-F OX and ABF1–4-R OX primers, Table S1 in File S1) into the pPS1 binary vector. The MBP-ABF1–4 fusion proteins for the EMS experiments were created by cloning the ABF1–4 PCR products (ABF1–4-F and ABF1–4-R primers; Table S1 in File S1) into the pMA-c2XL vector harbouring the maltose binding protein as tag for protein purification. Binary vectors were introduced into *Agrobacterium tumefaciens* LBA4404 by electroporation and used to transform *Arabidopsis* plants using the floral dip method [36].

Phylogenetic analysis

To identify members of the PLAT-plant-stress family from other plant species, multiple database searches were performed using the Basic Local Alignment Search Tool (BLAST) algorithms BLASTp and tBLASTn available on the public databases, PLAZA 2.0 (bioinformatics.psb.ugent.be/plaza) and Phytozome v8.0 (www.phytozome.net) with cutoff value of E<10⁻⁵. We used nucleotide and amino acid sequences of PLAT1 from the TAIR database (www.arabidopsis.org) to BLAST all databases. Phylogenetic analysis was performed by using CLUSTALW alignment in PHYLIP format clustal algorithm, and displayed in a phylogram tree format with locus name of each protein. Bootstrap values were presented as a percent of 100 resampled trees at each tree node using default settings of the TreeTop-Phylogenetic Tree (www. genebee.msu.su/services/phutreereduced.html).

Plant materials and growth conditions

*Nicotiana benthamiana* plants were grown under greenhouse conditions as described previously [37]. *Arabidopsis* plants (Col-0 ecotype) were grown in soil at 8 h light/16 h darkness at 22°C (light intensity: 180 μmol m⁻² s⁻¹) or on half strength MS medium under continuous light at 22°C (light intensity: 180 μmol m⁻² s⁻¹) in growth cabinets. T3 homozygous T-DNA insertion lines were obtained for the PLAT1 gene, plat1-1 SALK-112728c and plat1-2 SALK-1283454c, and the PLAT2 gene, platz SAIL-1171C06. The T-DNA insertions were verified with the primers PLAT1-1-F, PLAT1-1-R and SALK LB2 (plat1-1), PLAT1-2-F, PLAT1-1-R and SALK LB2 (plat1-2), and PLAT2-F, PLAT2-R and pROK2 LB1 (plat2) (Table S1 in File S1). For all plant experiments T3 or T4 homozygous plant lines were employed, based on the segregation of the respective antibiotic selection marker, except for the experiments shown in Figure S4, for which segregating T2 *Arabidopsis* lines were employed.

Abiotic stress conditions

The abiotic stress experiments in soil were performed as 3 biological replicates (cold stress as 2 biological replicates) with at least 10 plants each. For the *Arabidopsis* germination experiment, seeds were directly germinated on half strength MS medium including the respective chemicals as indicated. At least 100 seeds per treatment/genotype were used in 3 independent experiments. For the salt stress tolerance in plates, seeds were germinated and
grown for 6 d on half strength MS medium, transferred to half strength MS medium including NaCl (either 0, 150 or 200 mM) and optionally 5 µM dexamethasone (35S::>PLAT1 lines), and grown for another 4 d. These experiments were performed as 2 biological replicates, each with 2 technical replicates and with >12 Col-0 and >26 mutant/transgenics seedlings per plate. For the tunicamycin (TM) experiments, seeds were directly germinated on Col-0 and grown for another 4 d. These experiments were performed as 2 biological replicates, each with 2 technical replicates.

**Biotic stress conditions**

To determine pathogen susceptibility, leaves from 8-w-old Arabidopsis plants were infected with Pseudomonas syringae pv. tomato DC3000 with or without the avrRpm1 gene by infiltration using a needleless syringe as described previously [38]. Visual evaluation of disease symptoms were conducted at 3 to 5 d. For expression analysis, 8-w-old plants were infected with P. syringae pv. tomato DC3000 or Sclerotinia sclerotiorum. Subsequently, the S. sclerotiorum infected plants were kept in a clear plastic box under saturating humidity. The biotic stress experiments were performed as 3 biological replicates with at least 10 plants each.

**Expression analysis**

Total RNA isolation and Northern-blot analysis was carried out as described previously [37]. Filters were exposed to a screen for 4 d, which was scanned with a Phosphor imager (Fuji BAS2000, Ray-test, Germany). The probes for the PLAT1, PLAT2 and PLAT3 genes were generated by PCR from cDNA with the primers, PLAT1-F probe and PLAT1-R probe, PLAT2-F probe and PLAT2-R probe, and PLAT3-F probe and PLAT3-R probe (Table S1 in File S1), respectively. The RT-PCR analysis was performed essentially as described before [37]. At least 10 seedlings per genotype were grown for 14 d on control plates before transfer to the respective stress and control medium. The optimal cycle number was determined for each primer pair (Table S2 in File S1). Expression values were corrected for the ACTIN and UBQUITIN signal intensities and expressed as relative values compared to Col-0. For expression analysis, 3 independent experiments were performed.

**ABA determination**

The extraction and analysis of ABA was carried out as described previously [39].

**PLAT1-YFP localisation**

For transient PLAT1-YFP expression a single colony of A. tumefaciens LBA4404 containing either PLAT1:PLAT1-YFP or the ER-sh CD3-959 ER-marker [40] construct was inoculated into 5 ml induction medium with antibiotics and grown overnight at 28°C. The bacteria were collected by centrifugation and resuspended in 10 mM MES and 10 mM MgCl2 containing 200 µM acetosyringone to an OD600 of 1.0. Aliquots (1 ml) of A. tumefaciens cells containing PLAT1:PLAT1-YFP and ER-marker construct were mixed together, and then a syringe was used to infiltrate the mixture into the lower surface of N. benthamiana leaves. YFP and mCherry fluorescence was visualized 48 h post infiltration, using Olympus confocal laser scanning microscope (model FV10000, Tokyo, Japan). For stable PLAT1-YFP expression following ABA and salt treatment, the PLAT1:PLAT1-YFP line YFP13-1 was employed, 10 plants per treatment.

**Transactivation assay**

Suspensions of A. tumefaciens carrying the respective 35S:ABF1-1-4 overexpression constructs and the PLAT1:PLAT1-GUS rescue/reporter construct were mixed in a ratio of 1:1. The resulting mixed suspensions were used to infiltrate leaves of 6-w-old greenhouse grown N. benthamiana in soil. As control, leaves were infiltrated with A. tumefaciens carrying the PLAT1:PLAT1-GUS rescue construct only, or 10 mM MgCl2. The GUS fluorometric assays were carried out as described previously [41]. Samples were isolated from the infiltrated regions 2 d after infiltration for 5 independent plants and ground in 500 µl of extraction buffer containing 50 mM sodium phosphate, 10 mM EDTA, 10 mM β-mercaptoethanol and 0.1% N-lauroylsarcosine (pH = 7.4). After centrifugation at 4°C for 10 min at 13,000 rpm, the supernatant was used for the determination of GUS enzyme activity, 50 µl of supernatants was transferred into one slot of a black 96-well plate and 50 µl of a 2 mM MUG (methylumbelliferyl-β-D-gluconoride, Sigma) solution was added. For each sample 3 technical replicates were measured. The samples were incubated at 37°C for 30, 60 and 90 min, before the reaction was stopped with 1 M sodium carbonate. A standard curve was prepared with MUG (4-methylumbelliferyl, Sigma) in a concentration range from 0 to 16 µM. Excitation was measured at 365 nm, emission at 455 nm. Total protein amount was determined by the Bradford assay. GUS enzyme activity was calculated in pmol Mu min⁻¹ mg⁻¹ protein.

**Expression and purification of ABFs**

The ABFs were prepared employing a MBP-fusion purification procedure. Five ml of an overnight bacterial culture was incubated with 500 ml rich broth medium containing glucose and ampicillin. The cells were grown to 2x10⁶ cells ml⁻¹ (A₆₀₀ = 0.5). IPTG was added to a final concentration of 0.3 mM and a further incubation at 37°C for 2 h followed. The cells were harvested by centrifugation at 4,000 g for 20 min. The supernatant was discarded and the pellet re-suspended in 25 ml of column buffer (20 mM Tris-HCl, 200 mM NaCl, 1 mM EDTA, 1 mM DTT and 0.1 mM PMSF (pH = 7.4). The pellet was kept at −20°C overnight and thawed in cold water the next morning. The sample was placed in an ice-water bath and sonicated in short pulses of 15 s for at least 2 min. The suspension was centrifuged at 9,000 g for 30 min and the supernatant diluted 1/3 (v/v) with column buffer before purification. Amylase resin was poured in a 5 ml column and washed with 8 column volumes of column buffer. The diluted sample was loaded and slowly run over the column. The column was then washed with 12 volumes of column buffer and the proteins subsequently eluted with column buffer containing 10 mM maltose. 10 to 15 fractions containing 2 ml each were collected. Proteins were checked via SDS-page.

**Electrophoretic mobility shift assay**

For EMSA, the 200 bp PLAT1 promoter, generated by PCR with the primers pPLAT1-F and pPLAT1-R (Table S1 in File S1) was used as positive probe. The mutated version lacking the 2 ABRE elements was generated in 2 steps by PCR using the primers WIP-F1 and WIP-R1, and WIP-F2 and WIP-R2. The 2 PCR fragments were joined to create the negative 200 bp probe PLAT1 promoter fragment was labelled with [γ-³²P]ATP using T4 polynucleotide kinase (5’ end labelling). The reaction was incubated for 1 h at 37°C, purified and eluted with 10 mM Tris, pH = 8.0. The labelled probe was incubated 30 min at room temperature with 5 µg of the respective ABF protein extracts alone, with 100 fold molar excess of “cold” specific competitor (200 bp PLAT1 promoter), and with 100 fold molar
excess of “cold” negative probe (WIP1), including poly-dIdC as nonspecific competitor. The EMSA samples were run on a 5% native poly-acrylamide gel (10×10 cm). Before loading the samples, the gel was pre-run 40 min at 80 V and 4°C and the samples were run at 120 V and 4°C. After electrophoresis, radioactivity was detected in the dried gel as described above. Functionality of the EMSA assay and ABFx preparation using the MBP fusion protein was proven using the ABF1 protein extract and the published [34] positive ABRE-F and ABRE-R, and negative mABRE-F and mABRE-R control primers (Table S1 in File S1).

Statistical analysis
Standard deviations and average values were calculated in excel. Statistical significance for differences between treatments was analysed using the unpaired two sided Student’s t-test in excel. *** or * indicate statistical significance at p<0.001, p<0.01 or p<0.05, respectively.

Accession numbers
The AGI locus identifiers for the Arabidopsis PLAT-plant-stress family members are: PLAT1, At4G39750; PLAT2, At2G22170 and PLAT3, At5G65158.

Results

PLAT1 expression is induced by abiotic stress conditions

The PLAT-plant-stress subgroup of PLAT domain proteins comprises three Arabidopsis family members. Phylogenetic analysis of these Arabidopsis PLAT-plant-stress proteins using the neighbour-joining method showed that PLAT3 falls outside the other members (Figure S1A). In silico expression analysis by the eFP Browser [42] showed PLAT1 and PLAT2 expression during many developmental stages, which is affected under different stress conditions, whereas PLAT3 is not represented on the Affymetrix ATH1 arrays. To corroborate these data, we investigated expression of the PLAT family members by northern blot and RT-PCR analysis. This showed that PLAT1 was indeed expressed throughout development (Figure S1B). In contrast, PLAT2 expression was only detected in young seedlings (Figure S1E), whereas PLAT3 expression could not be detected in any of the analysed organs and developmental stages, neither by northern blot nor RT-PCR analysis at 35 cycles. PLAT1 expression was induced both by salt, following one day of watering with 200 mM NaCl (Figure S1C), and cold treatment, following incubation of 3-w-old plants at 8°C for 2 d (Figure S1D). PLAT1 expression was also induced following the transfer of young seedlings to medium with 200 mM NaCl (Figure 1). However, both the extent of PLAT1 induction and the temporal dynamics were different from that following salt stress using older plants grown in soil (Figure S1C), probably because PLAT1 expression is highest in young seedlings (Figure S1B). PLAT2 expression was repressed following the transfer of young seedlings to medium with 200 mM NaCl (Figure 1). These results confirmed the in silico expression data and suggested that among the Arabidopsis PLAT family members mainly PLAT1 is involved in abiotic stress responses.

PLAT1 promotes tolerance towards abiotic stress conditions

To analyse PLAT function in stress responses and plant development, we obtained T-DNA insertion mutants for PLAT1 and PLAT2. Two independent homozygous Arabidopsis loss-of-function mutants were obtained for PLAT1 (plat1-1 and plat1-2) and one promoter insertion mutant was obtained for PLAT2 (plat2) from the SALK and SAIL mutant collections, respectively [43,44]. Since both plat1 mutants exhibited similar phenotypic defects, only the characterisation of the plat1-1 mutant is described in detail. The plat1-1 and plat2 mutants exhibited no obvious growth defects under control conditions and most likely represent null alleles because PLAT1 and PLAT2 expression was not detected in the respective insertion mutants (Figure S1E).

The plat1-1 mutant was more sensitive to salt and drought stress, as well as cold stress conditions (Figure 2), evident by a reduction in root length upon growth at 8°C from 9.41±1.25 mm for Col-0 to 7.02±1.27 mm or 6.20±0.94 mm for plat1-1 and plat1-2, respectively (p<0.001, n = 14). In contrast, the plat2 mutant did not show obvious changes in salt stress tolerance (Figure 3). Together with the differential effect of salt stress on expression of the PLAT family members (Figure 1), these data support that only PLAT1 is involved in abiotic stress tolerance.

To analyse whether PLAT1 also plays a role in biotic stress responses, we studied PLAT1 expression following inoculation of plants with the hemibiotrophic pathogen P. syringae pv. tomato DC3000 or DC3000 avrRpm1 (RPM1, Figure S2A), and the necrotrophic fungal pathogen S. sclerotiorum (Figure S2B). This showed that PLAT1 expression was not specifically affected by these pathogens since increased expression was also observed for the respective control treatments (10 mM MgCl2 and PDA medium). Next, disease symptom development following inoculation with P. syringae pv. tomato DC3000 was investigated. Disease

![Figure 1. PLAT1 expression is induced by salt stress conditions.](https://example.com/figure1)

![Figure 2. PLAT1 loss-of-function reduces abiotic stress tolerance.](https://example.com/figure2)
confirmed that the defects in abiotic stress tolerance were indeed caused by the T-DNA insertion in the \textit{plat1-1} mutant and that the \textit{PLAT1} fusion proteins were functional in these rescued lines. PLAT1:PLAT1-YFP constructs: 5 out of 6 lines for PLAT1:PLAT1-GUS (Figure S4B). This confirmed that the defects in abiotic stress tolerance were indeed caused by the T-DNA insertion in the \textit{plat1-1} mutant and that the \textit{PLAT1} fusion proteins are functional in these rescued lines. Transgenic expression levels can be influenced to a large extent by plant sequences flanking the respective T-DNA integration sites, causing variation between individual transgenic lines harbouring the same T-DNA construct. In addition to transformed \textit{plat1-1} lines with restored (wild-type) ABA sensitivity, also transgenics were obtained exhibiting increased ABA sensitivity and thus tissue specific \textit{PLAT1} overexpression phenotype (Figure S4). To study whether \textit{PLAT1} overexpression would confer an opposite phenotype compared to the \textit{plat1} mutant, we selected these lines with tissue specific \textit{PLAT1} overexpression for further analysis and also generated ectopic \textit{PLAT1} overexpression lines. Because we could not obtain transgenics harbouring constitutive \textit{PLAT1} overexpression, we generated lines with inducible \textit{PLAT1} overexpression (35S::\textit{PLAT1}) employing the dexamethasone inducible OPELGR two component system [46]. From 17 transgenic \textit{PLAT1} OX lines, the majority exhibited delayed germination on 1.5 μM ABA (Figure S4C).

To compare tissue specific with ectopic overexpression, which could cause considerable differences in overexpression phenotypes [47], we selected the GUS1-3, GUS2-1, GUS3-5, YFP5-1 and YFP13-1 genomic rescue lines with increased ABA sensitivity in the seed germination assays for further analysis. In these selected tissue specific (GUS and YFP) \textit{PLAT1} overexpression lines, \textit{PLAT1} transcript levels were increased to a similar extend as the ectopic (OX) overexpression lines, compared to wild-type control (Table S4 in File S1). The different types of \textit{PLAT1} overexpression lines promoted salt stress tolerance after transfer to medium containing both 200 mM NaCl and 5 μM dexamethasone (DEX) (Figure 3). Thus, \textit{PLAT1} overexpression resulted in higher \textit{PLAT1} expression levels and reciprocal phenotypic effects compared to the \textit{plat1-1} mutant, confirming that \textit{PLAT1} functions in promoting tolerance towards abiotic stress conditions.

\textbf{PLAT1 promotes growth} The DEX induced \textit{PLAT1} OX lines from the salt stress experiment shown in Figure 3, appeared to show faster development on control medium including DEX (Figure 5A-E). Therefore, shoot and root growth was analysed in the \textit{plat1-1} mutant and \textit{PLAT1} overexpression lines. This showed that the different \textit{PLAT1} overexpression lines produced more shoot biomass compared to wild-type control (Figure 5F), while the length of the root apical meristem and total root length were not affected (Table S5 in File S1). Interestingly, the number of emerged lateral roots was increased, but only for the ectopic \textit{PLAT1} (OX) overexpression lines (Table S5 in File S1). This effect on lateral root formation was the only difference observed between the tissue specific and ectopic overexpression lines, and could reflect the difference between increased expression at its natural expression site (GUS and YFP) and broader expression in new (ectopic) cell types (OX). In contrast, \textit{PLAT1} loss-of-function did not affect either shoot or root growth, except for a reduced number of emerged lateral roots (Figure 5F, Table S5 in File S1).

\textbf{PLAT1 is expressed in structures related to the regulation of water household} The \textit{PLAT1:PLAT1-GUS} rescue/reporter construct conferred GUS activity (4 out of 6 lines analysed) in vascular tissue, leaf edges, hydathodes and stomata, which are structures related to the regulation of water household. The expression in stomata corresponds to the identification of \textit{PLAT1} in guard cells [48]. GUS activity could also be observed in floral organs, root tips, pericycle cells and lateral root primordia (Figure 6A-6H). The basal \textit{PLAT1} expression pattern in the shoot correlated with the abiotic stress tolerance, while that of the root correlated with the changes in lateral root number for the \textit{plat1-1} mutant and \textit{PLAT1} overexpression lines. Both treatment with salt and ABA resulted in

\begin{figure}
\centering
\includegraphics[width=\columnwidth]{figure3.png}
\caption{PLAT1 promotes tolerance towards salt stress conditions. Survival, expressed as the percentage of the seedlings transferred at the 6-d-old stage that developed (pale) green leaves during 4 d of salt stress conditions for the different mutant and transgenic lines (grey bars, n=26 per replicate) compared to Col-0 control plants (black bars, n=12 per replicate) that were grown on the same plates. Values are means of 3 replicates ± standard deviation. ** or * indicate statistical significance calculated using the unpaired Student's t-test at p<0.001, p<0.01 or p<0.05, respectively. doi:10.1371/journal.pone.0112946.g003}
\end{figure}
an expanded PLAT1 expression domain in root tips (Figure 6L-6N) and leaves (Figure 6O-6U), evident by the GUS activity in leaf mesophyll cells (Figure 6T and 6U), as well as in root pericycle cells (Figure S5).

To understand protein function, knowledge on its sub-cellular localisation is essential, because this influences access to and availability of interaction partners [49]. According to the Aramemnon membrane protein database [50], PLAT1 was predicted to contain a signal peptide involved in the secretory pathway and one transmembrane spanning domain. To analyse the sub-cellular localisation we transiently transformed the PLAT1:PLAT1-YFP construct together with different organelle markers [40] to N. benthamiana, which showed that the PLAT1-YFP fusion protein co-localised with the mCherry ER-marker ER-rk CD3-959 (Figure 7A-7C). The analysis of YFP reporter activity in the stable rescued plat1-1 transformants harbouring the PLAT1:PLAT1-YFP construct reflected the expression pattern evident from the GUS rescue reporter lines, confirmed PLAT1 induction by salt and ABA (Figure 7D-7I), and PLAT1 localisation to the ER (Figure 7J-7M). Further, PLAT1 is localised to rod shaped ER structures that resemble ER bodies (Figure 7M).

The PLAT1 promoter is a direct target of the ABF transcription factors

Sequence analysis of the PLAT1 promoter identified 2 G-boxes/ABRE elements (CACGTG motif) located at positions 2165 to 2156 and 2134 to 2126 relative to the transcription start site. ABRE elements are direct binding sites of the ABF/AREB (bZIP) transcription factors, which are positive mediators of the ABA signalling pathway [34,35]. We employed a transactivation assay to investigate whether the PLAT1 promoter is indeed regulated by these ABF transcription factors. Leaves from wild-type tobacco N. benthamiana plants were simultaneously infiltrated with A. tumefaciens harbouring either one of the 35S:ABF1–4 overexpression constructs (ABF1–4) and the PLAT1:PLAT1-GUS genomic rescue/reporter construct (pGUS). Each of the double infiltrations resulted in significantly higher GUS activity levels, compared to infiltration with the PLAT1:PLAT1-GUS reporter construct only (Figure 8). This indicated that the ABF transcription factors activate the PLAT1 promoter.

Electrophoretic mobility shift assays (EMSA) confirmed these transactivation results and showed that the ABF1, ABF3 and ABF4 transcription factors directly bind to the PLAT1 promoter fragment (~200 to +1), which contains the 2 ABRE elements. This ABF binding was specifically competed with a 100-fold molar excess of unlabelled 200 bp PLAT1 promoter fragment (PLAT1; Figure S6), but not with the mutated 200 bp promoter fragment WIP1, where point mutations were introduced in the

Figure 4. PLAT1 loss-of-function reduces ABA sensitivity during seed germination. (A) Seed germination of plat1-1 and wild-type (Col-0). (B) Seed germination of plat1-1 and wild-type on 300 mM mannitol (osmotic stress). (C) Seed germination of plat1-1 and wild-type on 200 mM NaCl (salt stress). (D) Seed germination of plat1-1 and wild-type on 1.5 µM ABA. Values are means of 3 replicates ± standard deviation. n=100 per replicate.
doi:10.1371/journal.pone.0112946.g004

Figure 5. PLAT1 promotes plant growth. Phenotypes of plants from control medium including 5 µM DEX from the salt stress experiment shown in Figure 3. (A) Wild-type. (B) plat1-1. (C-E) Three independent PLAT1 ectopic overexpression lines (OX). Scale bar = 1 cm. (F) Shoot biomass production (weight per 5 shoots) for the different PLAT1 lines. Values are means of 8 replicates ± standard deviation. ** or * indicate statistical significance calculated using the unpaired Student’s t-test at p<0.001, p<0.01 or p<0.05, respectively.
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two ABRE elements (WIP1; Figure S6). Thus, PLAT1 expression is regulated by the ABA signalling pathway as direct target of the ABF transcription factors.

PLAT1 promotes tolerance towards ER stress elicited by tunicamycin

The PLAT1 subcellular localisation in the ER (Figure 7) suggested that PLAT1 function is related to ER stress. Under stress conditions, misfolded proteins accumulate in the ER that eventually cause ER stress [51], resulting in reduced growth and induction of the unfolded protein response (UPR) to compensate for this increased accumulation of misfolded proteins. TM is used as ER stress agent and interferes with N-linked glycosylation of secreted glycoproteins, which prevents protein folding in the ER [51]. The tolerance of the different PLAT1 overexpression lines towards TM was improved compared to Col-0 wild-type (Figure 9). In contrast, the plat1-1 mutant was more sensitive, while the response towards TM was not affected in the plat2 mutant (Figure 9), which supports a function for PLAT1 in ER stress responses and/or UPR. The basal expression levels for the ER stress markers BIP1,2, CNX1, CRT1 and PDIL [52] was higher for the different PLAT1 overexpression lines (Figure S7), which suggests that the capacity of these lines to deal with unprocessed and/or misfolded proteins in the ER is increased, which could contribute to an increased abiotic stress tolerance.

Discussion

The complete Arabidopsis genome sequence is available, nevertheless only for a relatively low percentage of Arabidopsis proteins experimental evidence concerning their function is
available. While among the PLAT-plant-stress family only CgTIN1 function has been studied by heterologous expression in tobacco [4], for other family members only limited in silico expression data is available. This showed that these members were induced by different abiotic and biotic stimuli, suggesting that PLAT-plant-stress proteins in general could promote tolerance towards stress responses and thus can be of great importance for developing stress tolerant crops. Nevertheless, little information is available on the function of these proteins. We assigned a function to the previously uncharacterised Arabidopsis PLAT domain proteins, which are members of this PLAT-plant-stress subgroup. The combination of genetic and physiological approaches supported functional diversification within this Arabidopsis PLAT protein family. PLAT1 critically functions as positive regulator of abiotic stress tolerance and also confers increased plant growth. Because PLAT1 is a direct activated target of the ABF transcription factors and PLAT1 levels affect ABA sensitivity based on seed germination assays, PLAT1 represents a novel downstream target of the ABA signalling pathway. PLAT2 appears to function specifically in seed dormancy, while PLAT3 might represent a non-expressed pseudogene.

Analysis of the plat1-1 loss-of-function mutant, PLAT1 overexpression lines and genetic complementation of the plat1-1 mutant, showed that PLAT1 critically functions as positive regulator of abiotic stress tolerance. Abiotic stress treatments induced PLAT1 expression, PLAT1 loss-of-function resulted in reduced abiotic stress tolerance, whereas PLAT1 overexpression conferred an inverse response, evident from increased cold, drought and salt stress tolerance. No change in pathogen susceptibility was detected for the plat1-1 loss-of-function mutant, which correlates with the fact that pathogen infection did not significantly affect PLAT1 expression. It is however possible that an altered pathogen resistance in plat1-1 could have been too weak to be detected in the employed pathosystem. Otherwise, functional redundancy among the PLAT family members (partially) could have compensated for PLAT1 loss-of-function in biotic stress tolerance.

High ABA levels confer resistance towards abiotic stress conditions like drought and salinity, which is linked to its role in regulating stomatal aperture, while ABA shows antagonistic interaction with pathogen defence signalling pathways [26,28–31]. The ABF/ABRE transcription factors, which are positive mediators of the ABA signalling pathway [34,35], were shown to be induced by cold (ABF1), salinity (ABF2 and ABF3) and drought stress (ABF4) [34], while ABF2 or ABF3 overexpression enhanced abiotic stress tolerance [53]. PLAT1 overexpression increased and loss-of-function reduced sensitivity towards ABA during seed germination. Because ABA levels were not significantly affected in the different PLAT1 lines, PLAT1 overexpression resembled the effect of increased ABA signalling, including the differential regulation of abiotic and biotic stress tolerance.

Although it is widely speculated that the effect of ABA signalling on stress tolerance in higher plants is regulated by the complex (antagonistic) interactions with other phytohormones, our understanding of the ABA signalling pathway leading to the adaptation of naturally occurring multi-stress responses remains unclear [54]. Genes harbouring two ABRE elements in their promoter were predicted to be direct (activated) target genes of the ABF/ABRE transcription factors [34] and the expression for a large number of such genes was affected by a triple ABA loss-of-function mutant [35]. However, only the DREB2C and RD29B genes were functionally shown to be direct ABA target genes [55,56]. The PLAT1 promoter harbours two ABRE elements, located at positions −163 to −156 and −134 to −126 relative to the transcription start site. In agreement with these findings, transactivation and EMSA experiments showed that the ABF transcription factors directly bind to and activate the PLAT1 promoter. Analysis of plat1-1 lines complemented with the PLAT1:PLAT1-GUS rescue/reporter construct showed that PLAT1 is expressed in the leaf vasculature, hydathodes and stomata, which are structures linked to the regulation of water household, but also at specific regions in the root. Together with the induced PLAT1 expression levels as well as expanded expression domain following ABA treatment and abiotic stress conditions, PLAT1 expression correlates with PLAT1 function in abiotic stress tolerance and its regulation by ABA signalling. Together, these data showed that PLAT1 represents a novel component of the ABA signalling as direct target of the ABF transcription factors, which might explain its function in stress tolerance.

ABA in general negatively affects plant growth, mainly through crosstalk with the brassinosteroid pathway [57], and promotes quiescence of stem cells resulting in reduced root growth and
lateral root formation [58–60]. In addition to the effects on stress tolerance, which could be correlated with PLAT1 being a downstream target of the ABA signalling pathway, PLAT1 promotes plant growth, evident by a faster development and consequently increased shoot biomass. Therefore, PLAT1 function exhibits both expected and unexpected ABA related responses. Heterologous CaTIN1 overexpression in tobacco also resulted in increased abiotic stress tolerance and plant growth, but additionally conferred increased biotic resistance, probably through influencing the redox state. Further, CaTIN1 expression was induced by ethylene treatment and infection by tobacco mosaic virus, but not following ABA treatment [4]. Thus, despite the fact that PLAT1 and CaTIN1 are orthologs, partially convergent evolution occurred in the different plant species on protein function and transcriptional regulation.

Transient expression experiments and analysis of plat1-1 lines complemented with the PLAT1:PLAT1-YFP rescue/reporter construct showed that PLAT1 is localised to the ER, but also in rod shaped structures resembling ER bodies. This is supported by the induction of these structures with PLAT1-YFP signals following ABA treatment or salt stress conditions. ER bodies are specific to Brassicaceae and induced following stress conditions and wounding [61–63], but no direct correlation between ER bodies and abiotic stress responses has been shown. PLAT1 promotes tolerance towards the ER stress elicitor TM and the basal expression levels of ER stress markers, representing chaperonins functioning in ER stress and/or UPR.

Together our results indicate that PLAT1 functions in abiotic stress tolerance, either directly through promoting abiotic stress responses, and/or indirectly through improving basal tolerance/fitness. A direct promotion of abiotic stress responses could result from promoting ABA signaling, which is related to its function as novel direct target gene of the ABA signalling pathway. An indirect effect could result from stimulating ER stress responses for a higher basal tolerance/fitness. ER stress responses were shown to be indispensable for abiotic stress responses [64,65] and the ER appears to play a prominent role in ABA-mediated stress signalling since ABA release from the ER is important for plants coping with stress [66]. The PLAT1 protein essentially harbours one transmembrane spanning domain and one large PLAT domain that covers the rest of the protein sequence and which has been shown to function in protein interaction. Therefore, PLAT1 most likely does not possess enzymatic activity, but rather functions as ‘docking site’ for interacting proteins with enzymatic or signalling activity functioning in ABA regulated pathways, enabling PLAT1 to regulate their activity.

To obtain plants through biotechnology or breeding approaches with increased tolerance towards adverse conditions, but without yield penalties under optimal growth conditions, it is important to identify all genes involved in stress responses and understand their function. Therefore, the identification and assignment of a function to the previously uncharacterised PLAT-plant-stress family member PLAT1 contributes to this important goal. The improved plant growth associated with the increased tolerance towards cold, drought and salt stress mediated by PLAT1 overexpression could be an important asset in crop improvement. To enable the application of PLAT1 or other members from the PLAT-plant-stress family in crop improvement, future studies will be needed to address the multifaceted role of these proteins in stress tolerance and plant development.

Supporting Information

Figure S1 PLAT1 expression patterns under different conditions. (A) Phylogenetic tree of the PLAT-plant-stress subgroup. Phylogenetic analysis was carried out using the neighbour-joining method with 100 bootstraps and displayed using TreeTop. Glycine max (Glyma), Zea mays (GRMZM), Oryza sativa (Os), Populus trichocarpa (POPTR), Sorghum bicolor (Sh). (B) PLAT1 expression in different organs from 3-w-old, 6-w-old and 12-w-old wild-type (Col-0) plants: F, Flower; H, hypocotyl; L, leaf; R, root; S, inflorescence stem and W, whole plant. (C) PLAT1 expression following salt treatment (right) compared to control watering (left). (D) PLAT1 expression following cold treatment. (E) PLAT1 and PLAT2 expression by RT-PCR in the respective T-DNA insertion mutants plat1-1 and plat2. Bottom panels, rRNA for loading control (A-D).

Figure S2 PLAT1 loss-of-function does not affect biotic stress tolerance. (A) PLAT1 expression following leaf infiltration of 10⁶ cfu ml⁻¹ of P. syringae pv. tomato DC3000 or DC3000 avrRpm1 in 10 mM MgCl₂ compared to control treatment (MgCl₂). (B) PLAT1 expression following infection with S. sclerotiorum compared to control treatment (PDA). (C) Leaves from wild-type (Col-0) plants (Top panel) and plat1-1 plants (Bottom panel), 3 d after infection with 10⁵ cfu ml⁻¹ P. syringae pv. tomato DC3000. Scale bar = 1 cm, n=10. (TIF)

Figure S3 PLAT2 functions in seed dormancy. Seed germination of plat1-1, PLAT1:PLAT1-GUS line GUS3-5, 35S::>PLAT1 line OX9-6, plat2 and wild-type (Col-0) on control medium including 3 µM DEX without prior stratification. Values are means of 3 replicates ± standard deviation. n=100 per replicate. (TIF)

Figure S4 Increased ABA sensitivity by tissue specific or ectopic PLAT1 overexpression. (A, B) Seed germination of plat1-1, wild-type (Col-0) and plat1-1 lines complemented with the PLAT1:PLAT1-GUS rescue construct (GUS) (A), or plat1-1 lines complemented with the PLAT1:PLAT1-YFP rescue construct (YFP) (B) on medium supplemented with 1.5 µM ABA. (C) Seed germination of wild-type and transgenic lines harbouring the 35S::>PLAT1 ectopic overexpression construct (OX) on medium supplemented with 1.5 µM ABA and 5 µM DEX. Values are means of 3 replicates ± standard deviation. n=100 per replicate. (TIF)

Figure S5 PLAT1 expression is induced in adult roots by ABA treatment and salt stress. PLAT1:PLAT1-GUS seedlings were monitored for PLAT1 expression 8 h (A-C) and 24 h (D-F) following transfer to control, NaCl or ABA plates. (A, D) Detail of 2-w-old adult root with PLAT1 expression in emerging lateral root primordia following transfer to control medium. (B, E) Detail of adult root with expanded expression domain following transfer to 200 mM NaCl. (C, F) Detail of adult root with expanded expression domain following transfer to 1.5 µM ABA. Scale bar = 0.1 mm, n=10. (TIF)

Figure S6 PLAT1 functions as direct ABF target in ABA signalling. EMSA assay showing that the ABF transcription factors bind to the 200 bp PLAT1 promoter region containing two ABRE elements PLAT1 (*). This binding was specifically competed with a 100 molar excess of unlabelled PLAT1 promoter fragment (PLAT1), but not the negative probe lacking the two
PLAT1 Regulates Abiotic Stress Tolerance

ABRE elements (WIP1). Arrowhead indicates shifted band. Bracket indicates free probe.

Figure 7 PLAT1 overexpression lines exhibit higher basal expression levels for ER stress markers. Relative expression levels for ER stress markers in the different PLAT1 overexpression lines compared to wild-type (Col-0) and the basal expression levels for ER stress markers. (A) BIP1.2 (HSP70), (B) CNX1 (CALnexin), (C) CRT1 (CALreticulin1) and (D) PD1 (PROTEIN DISULFIDE ISOMERASE-like). Values are means of 3 replicates ± standard deviation. n=10 per replicate. ***, ** or * indicate statistical significance calculated using the unpaired Student’s t-test at p<0.001, p<0.01 or p<0.05, respectively.

(TIF)

File S1 Combined file containing supporting tables. Table S1: List of primers used for cloning, genotyping of T-DNA insertion mutants and EMSA controls. Table S2: List of primers used for RT-PCR. Table S3: ABA levels are not affected by PLAT1. Table S4: PLAT1 transcript levels in the different overexpression lines. Table S5: PLAT1 promotes lateral root formation.

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Author Contributions

Conceived and designed the experiments: TKH AA DKG EvgG TR. Performed the experiments: TKH AA SHE DKG HB UJ YR WA EvgG. Analyzed the data: TKH AA DKG EvgG. Contributed reagents/materials/analysis tools: SK EvgG TR. Wrote the paper: TKH AA EvgG TR.

References


